# Schematics of the kinetoplastid mitochondrial kDNA organization









### And how were these networks labeled

In contrast to the ones before, these were labeled in vitro, after the networks were isolated.

DNA polymerase or Terminal deoxynucleotidal transferase (TdT) were added to the isolated networks. Both will be active on Nicked or Gapped substrates (DNA Polymerase can add Labeled nucleotides to the 3'OH and so can TdT.



Con LOA polymetries (Unite C) take, characteristing, while, the machines were conducted at  $10^{-1}$ , for  $0^{-1}$  and  $10^{-1}$  and  $10^{-1}$ , for  $0^{-1}$  and  $10^{-1}$  and  $10^{-1}$ 

Same experiment, different label

### kDNA rotates during replication





## ...except in T brucei, where it oscillates





#### How are these kDNA networks labeled?

Two methods: In vivo: PULSE/CHASE

In vitro: Labeling nicked and gapped Minicircles

In this figure, the in vivo labeling trick was used:

Cells are grown in the presence of a radioactive Thymidine for A) 3 min B) 6 min C) 30 min and D) 60 min (That's the Pulse). Any DNA replicated during the pulse will incorporate the radioactive Thymidine. A Pulse/chase experiment is when you grow the cells in the presence of the labeled substrate (pulse) and then grow the cells in the presence of unlabeled substrate for a defined period of time after (chase)



Figure 1. EM Autoradiographs of kDNA Networks Labeled In Vivo with [PH]Thymidine

Labeling periods were: (A), 3 min; (B), 6 min; (C), 30 min; (D), 60 min. Propidium diiodide, used to distinguish nicked or gapped minicircles from colvalently closed minicircles, was present in the hypophase and spreading solution at 10  $\mu$ g/ml. Autoradiographic exposure times were: (A and B), 11 weeks; (C and D), 40 hr. Bars, 3  $\mu$ m.